

# Ecoli NightSeq®

## Sample Preparation

### Input Material

#### • *E. coli* colony

Grow your *E. coli* on agar plates long enough to have a colony diameter of at least 1 mm. This will ensure a high cell density and is a prerequisite for reliable sequencing.

#### • 5 µl of a bacterial suspension

(e.g. LB liquid culture).

#### • 1 µl of plasmid DNA

(if concentrations are extremely low)

Besides *E. coli*, other Gram-negative bacteria can also be sequenced (e.g. heat killed *Pseudomonas aeruginosa*). Please ask for details.

### Procedure

1. Shortly centrifuge the tube/plate<sup>1</sup> to avoid having liquid on the lid/foil. Carefully remove the sealing foil from the plates.

2. Wear protective gloves and use a toothpick<sup>2</sup> to take as much of the *E. coli* colony as possible and inoculate into our tubes/plates. Swirl the toothpick for some seconds to transfer as many cells as possible into the liquid.

In case of bacterial suspension/plasmid DNA use a pipette to add the specific volumes.

**For Plates:** Please fill plates in **vertical order** (not horizontal!) and **avoid empty wells** between full wells.

3. **To make full use of the time advantage, create a replica right after:** Use the same toothpick to inoculate LB media containing antibiotics to produce an overnight culture (or streak out on a fresh agar plate). The provided additional sticker can be used to label your replica.

4. Login to our webshop at [www.microsynth.ch](http://www.microsynth.ch) to place your order for the Ecoli NightSeq® service.

5. Finally put your sample(s) into a transparent plastic bag and drop it into the closest sample drop box. **No incubation needed.**

6. Your sequencing results will be available the next working day before 2 pm for tubes and from 3 pm for plates.

<sup>1</sup> Tubes and plates are delivered together with the barcode labels.

<sup>2</sup> Or something similar that can be used for this purpose like a pipette tip, etc.

Simply stick a **red barcode label** on your sample tube/plate as shown in the image. Please do not put any sticker onto the lid and do not wrap it with parafilm!

**Remark:** Never pre-mix with the primer. Enclose a separate tube of specific primer (10 µM aliquots).



## Need More Information?

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